

# AIS\_quant manual

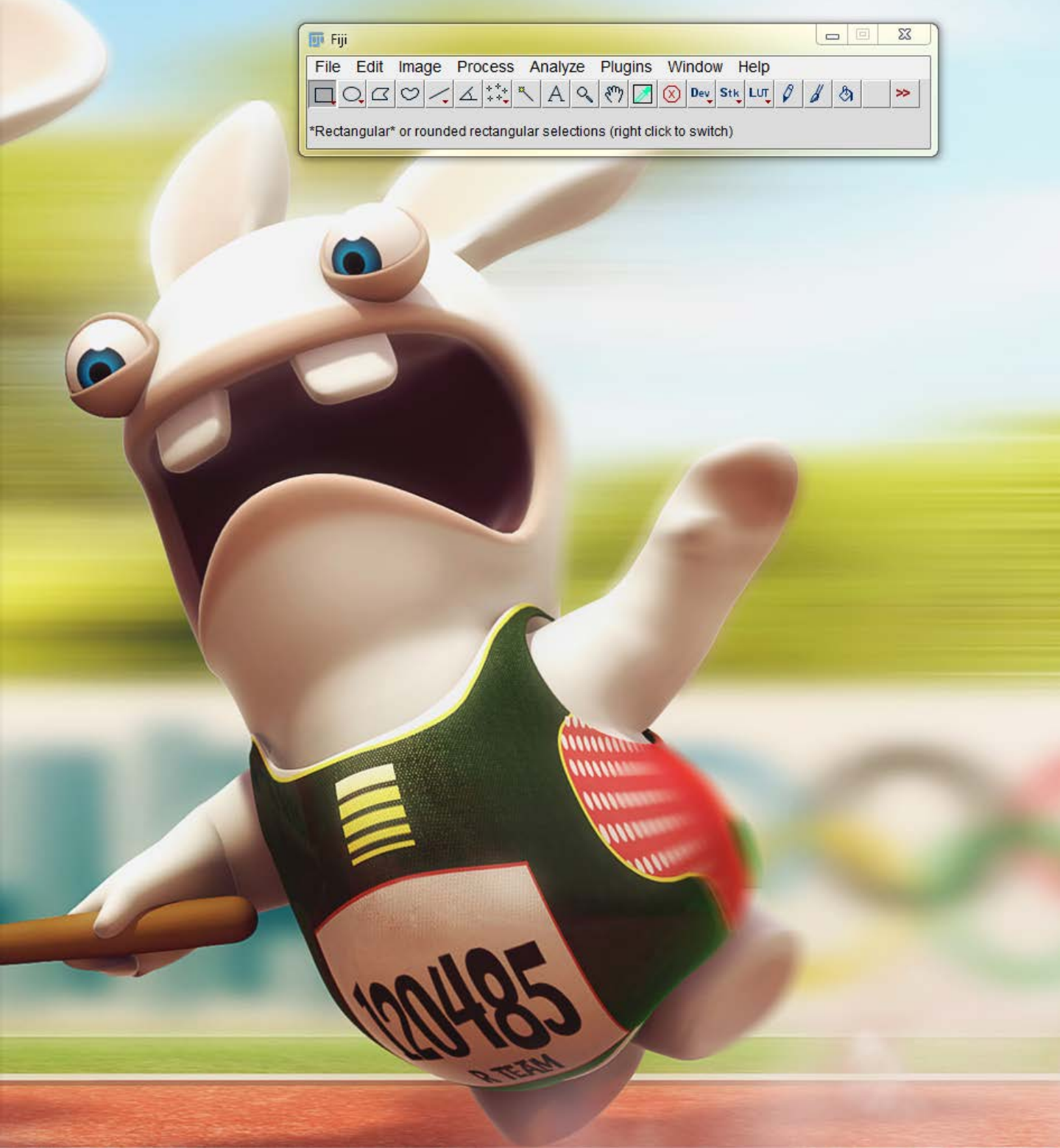
Version 1.0

2012-11-26

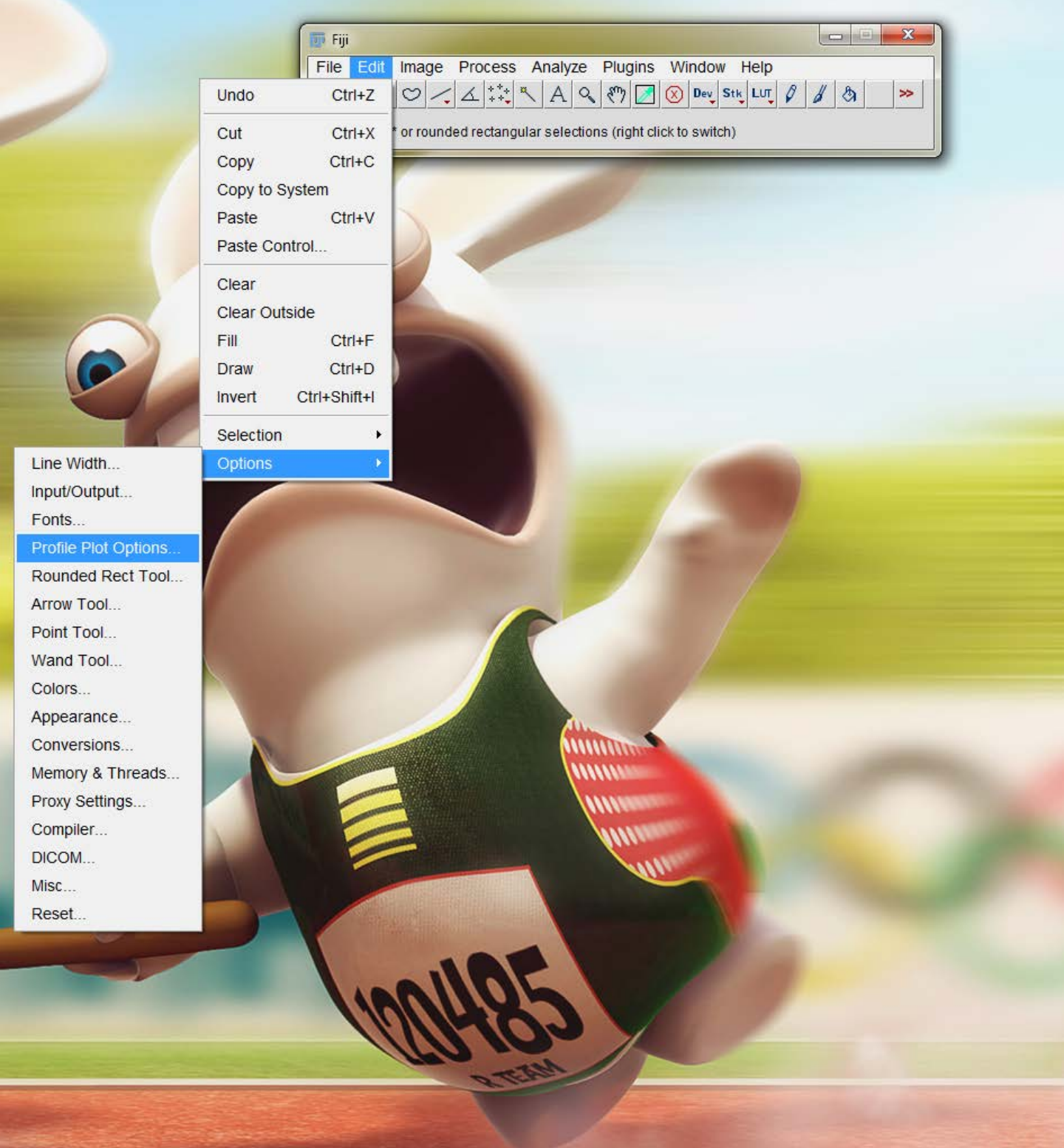
Sam van Beuningen

Please read before use: M. S. Grubb, and J. Burrone, 'Activity-Dependent Relocation of the Axon Initial Segment Fine-Tunes Neuronal Excitability', *Nature*, 465 (2010), 1070-4.

Many thanks to Eugene Katrukha

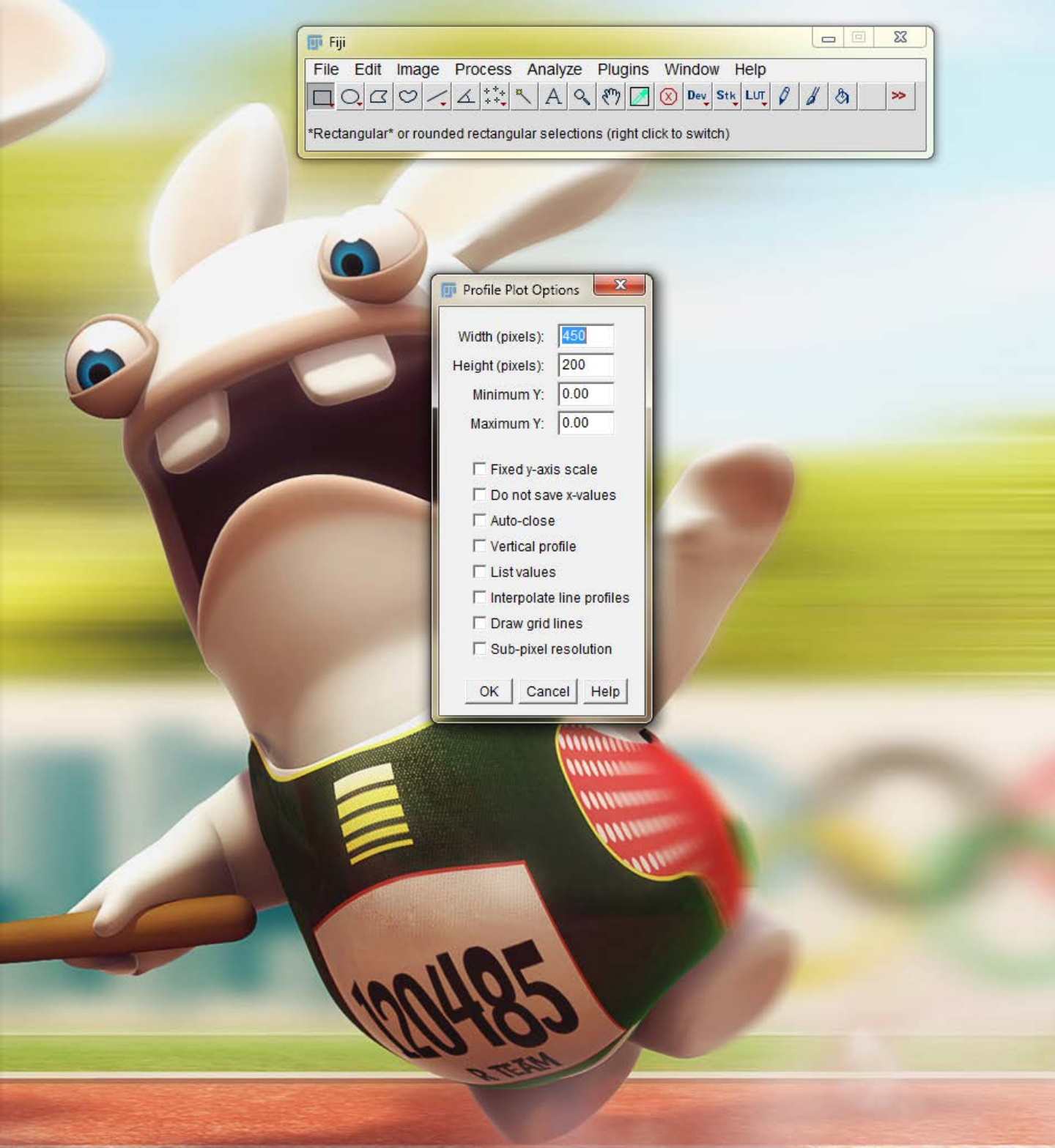


- Open ImageJ or Fiji

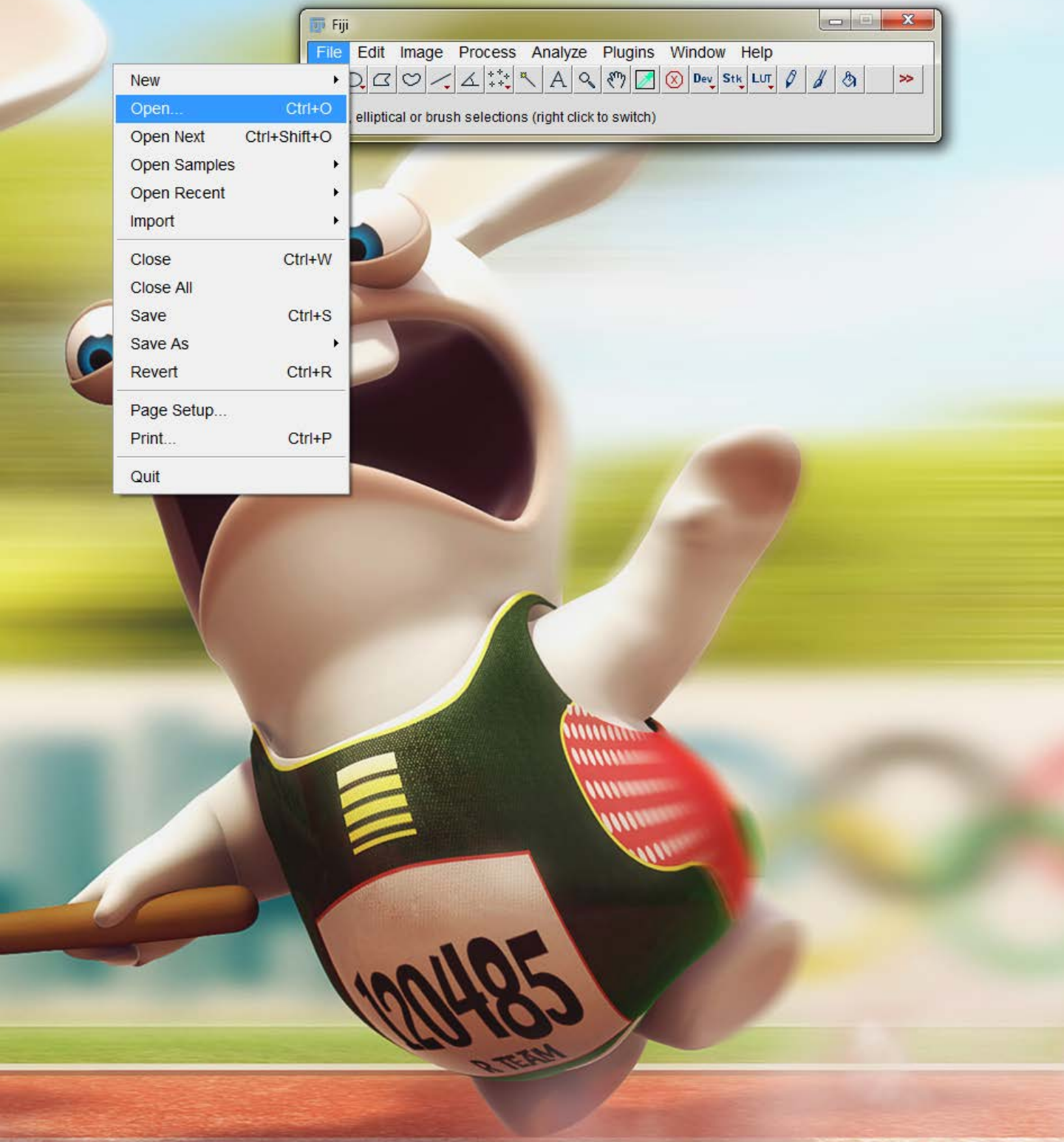


- Check Profile Plot Options

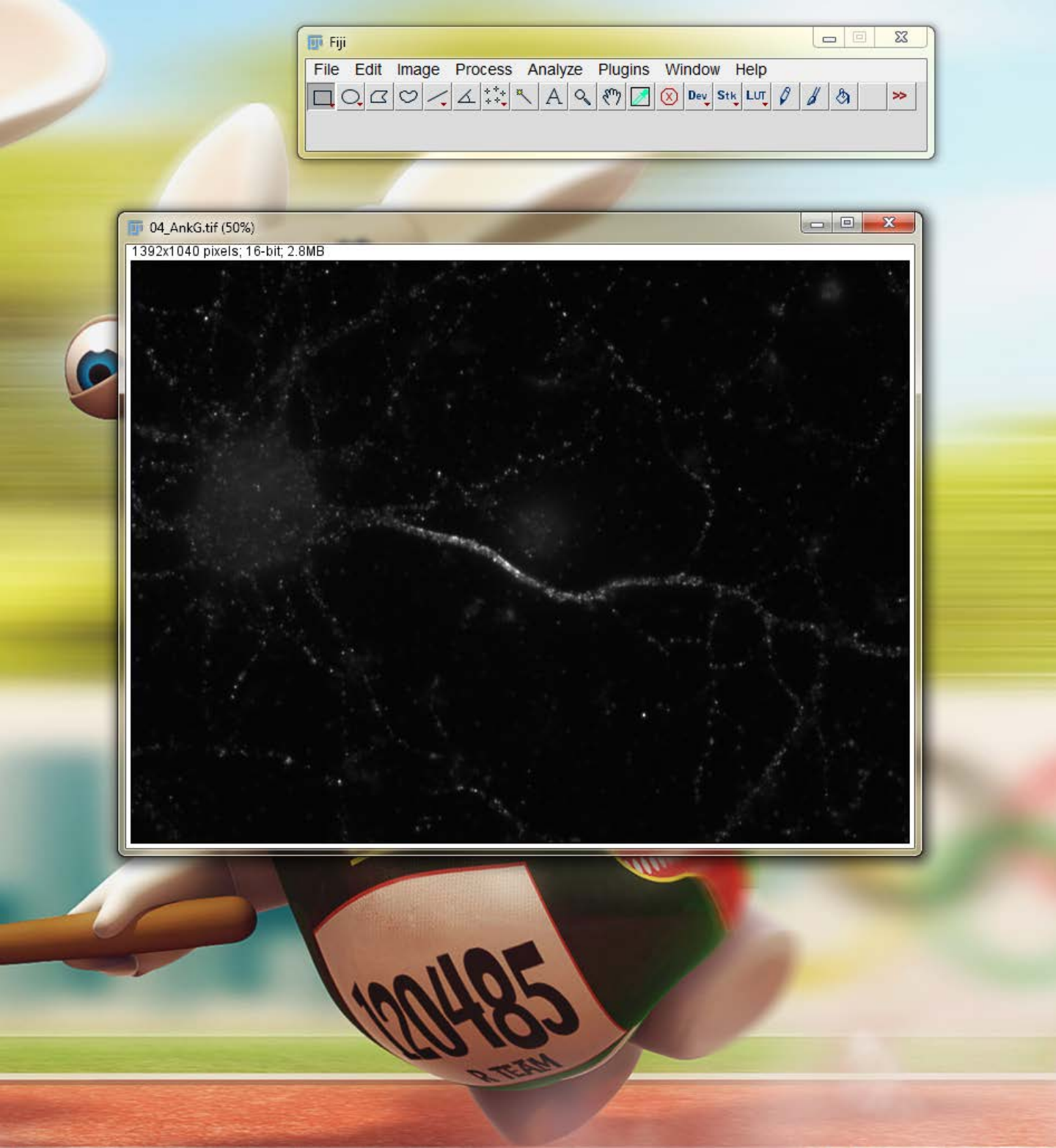




- Be sure that everything is unchecked

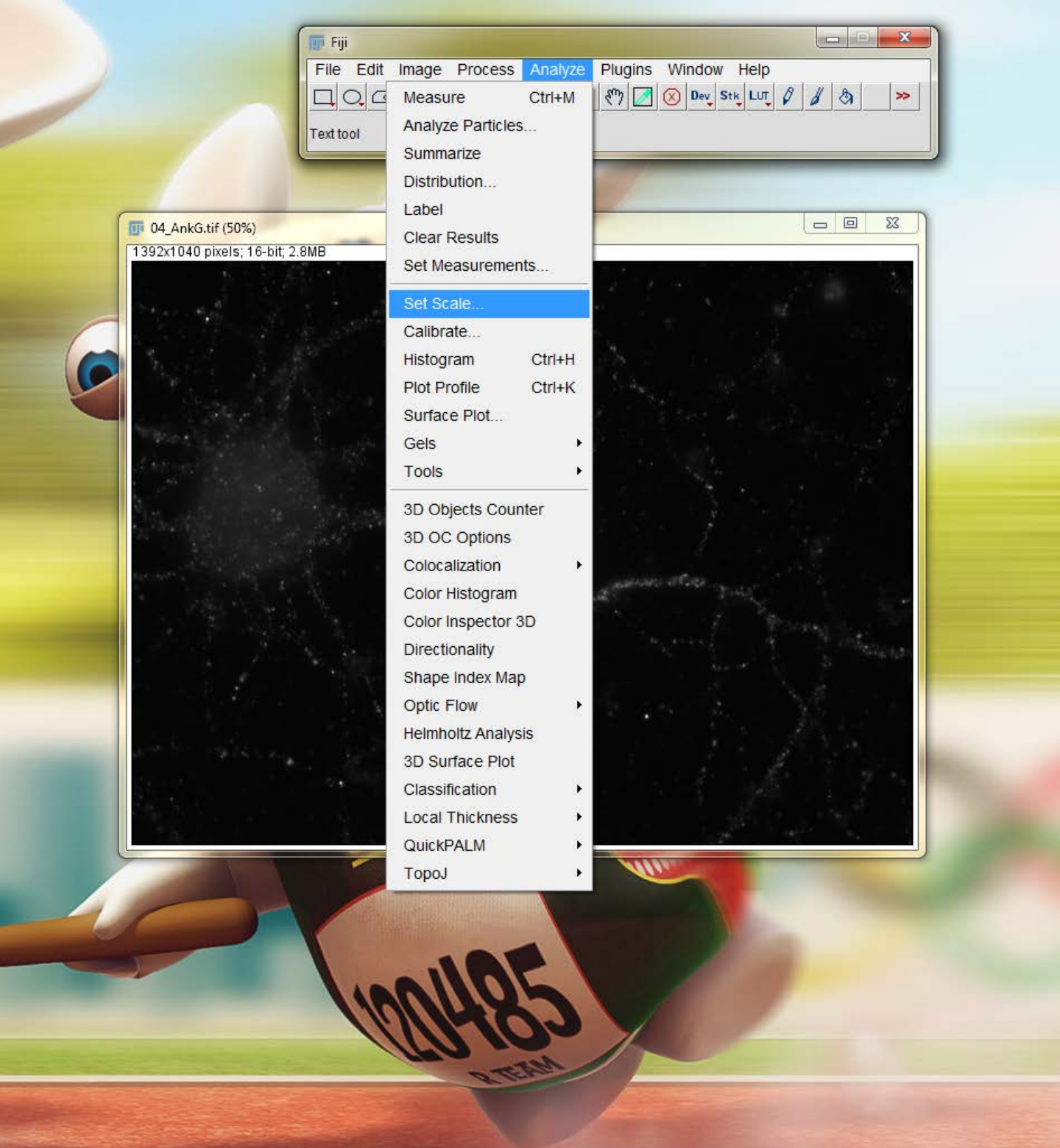


- Open a picture with your staining (you can also drag it into the program).

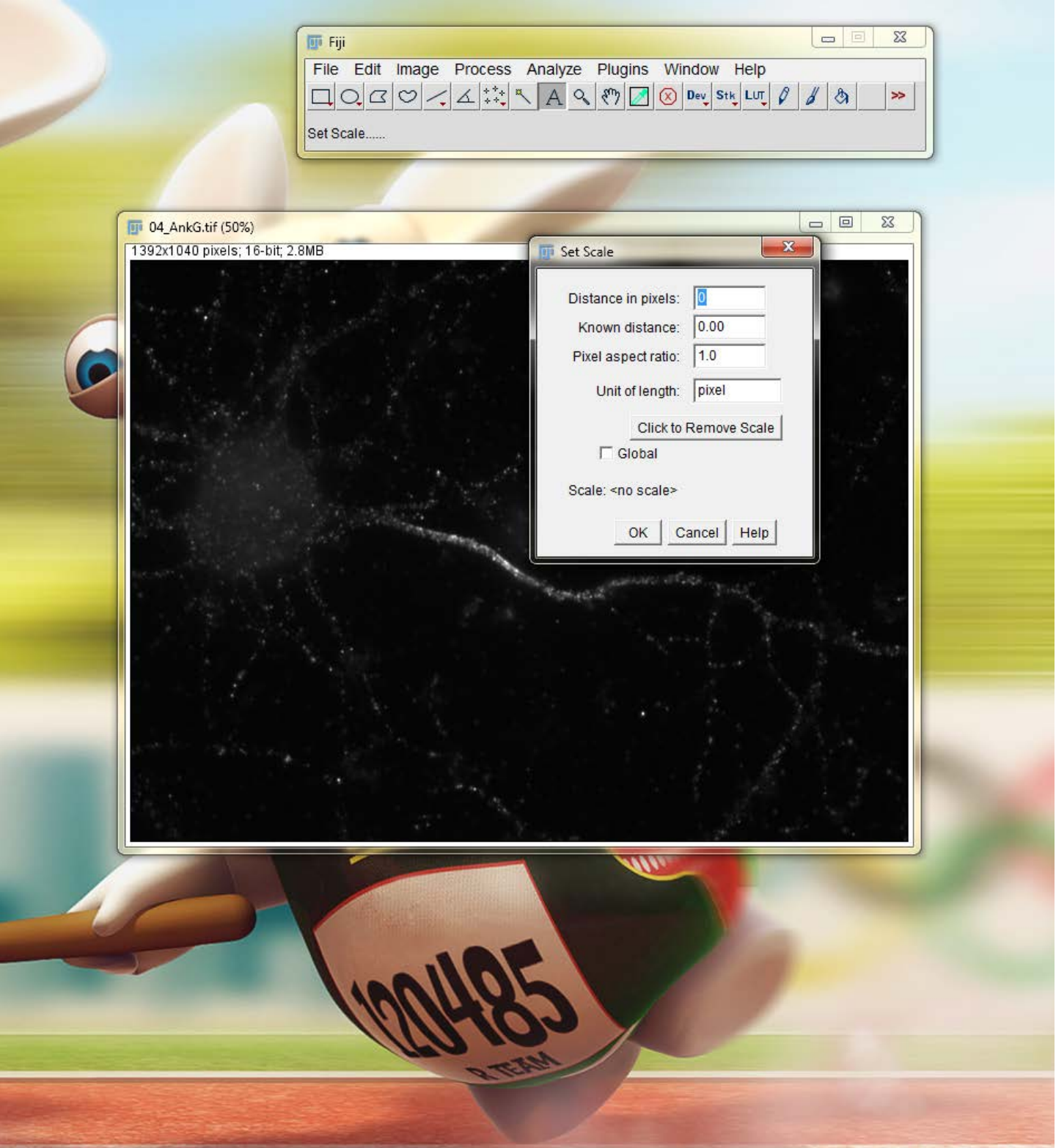


- Is that really the best staining you can get?
- Yes?
- Ok lets continue...



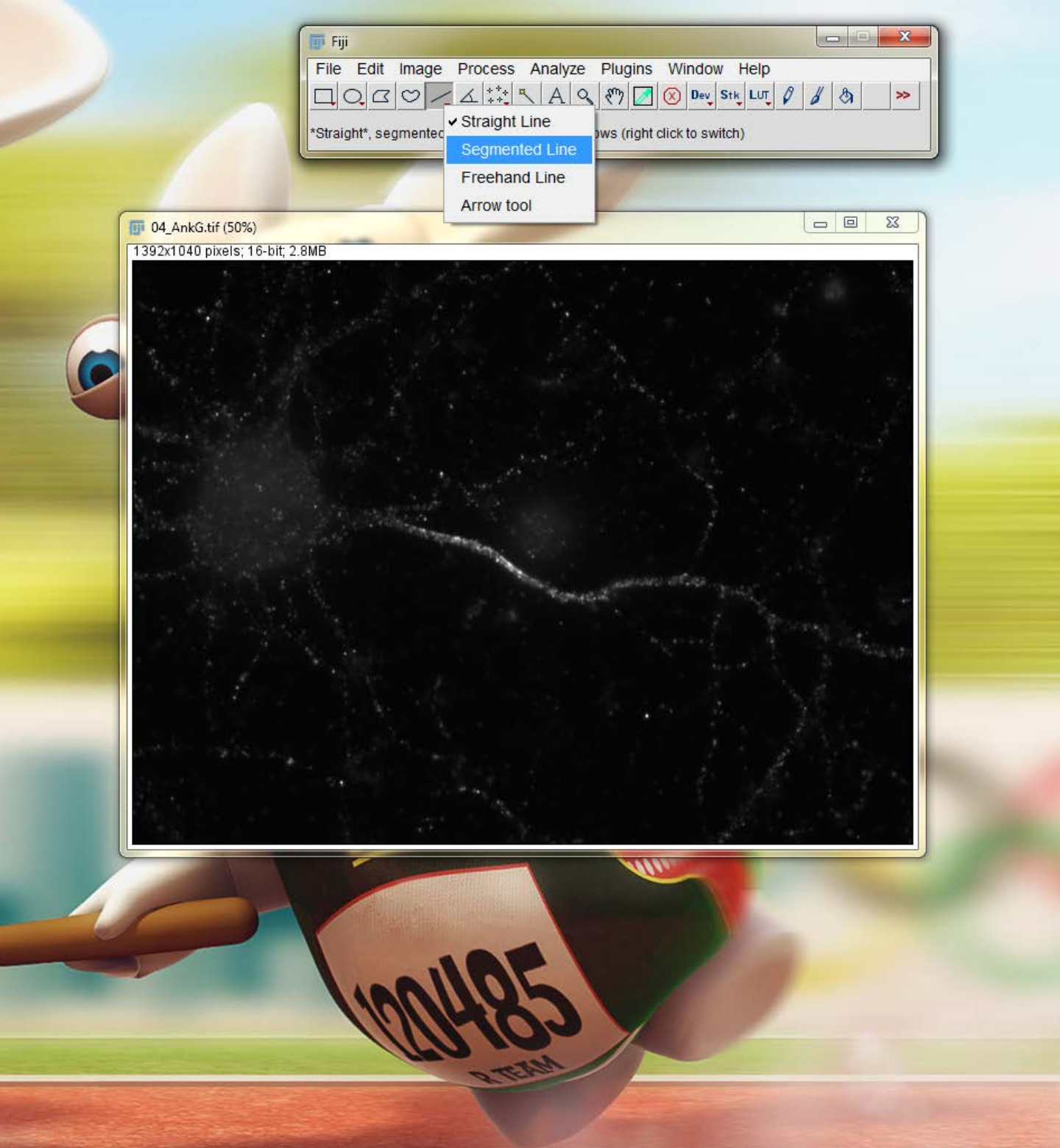


- Check the scale of your picture since some microscope already save the correct scale in the image.

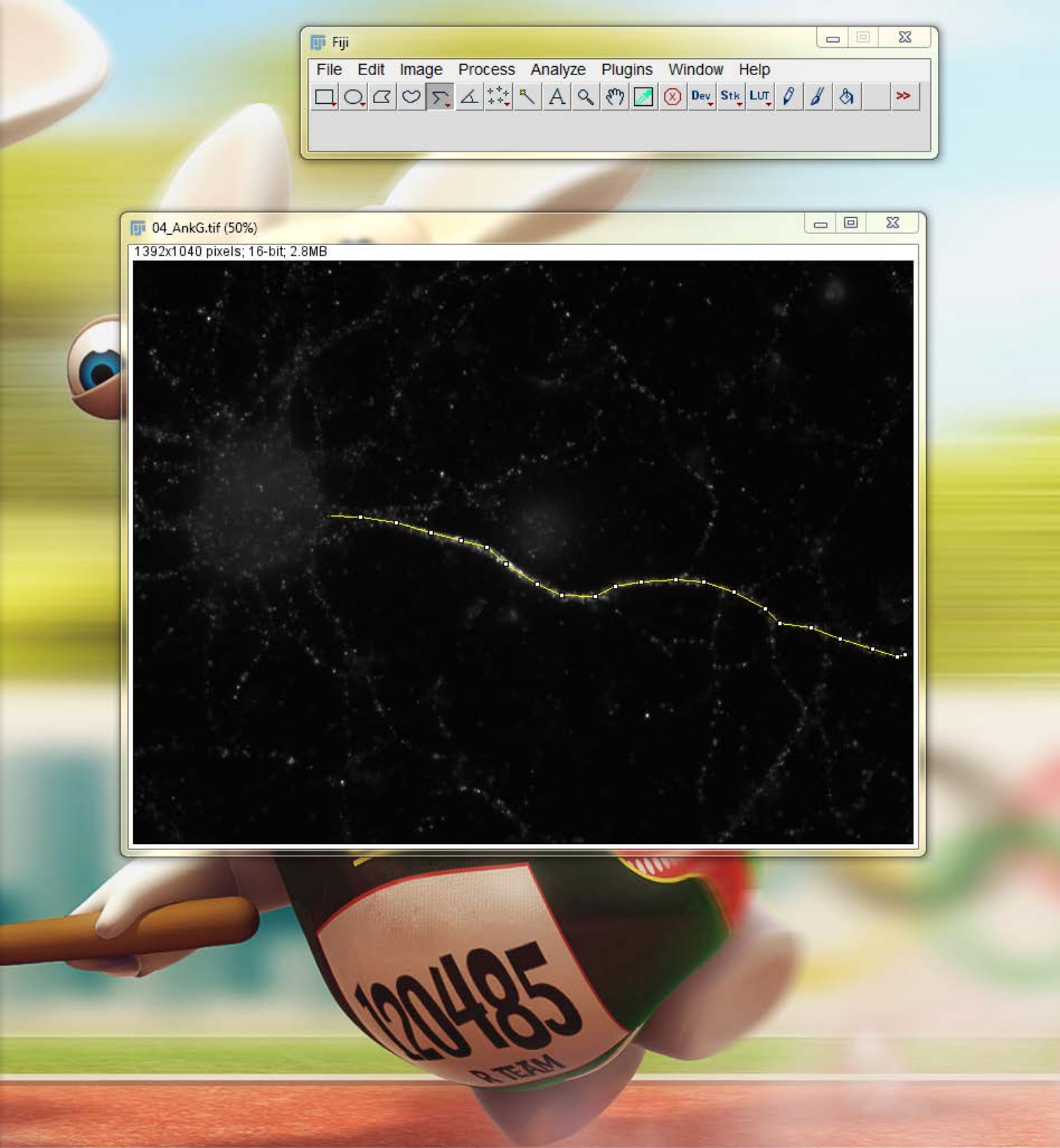


- Shown here is the standard scale of 1 pixel, which is good.
- Sometimes this scale is set to microns per pixel.
- If so, you could either remove the scale by using the button.
- Or you could leave it but then you should keep in mind that you already have the correct scale later on.

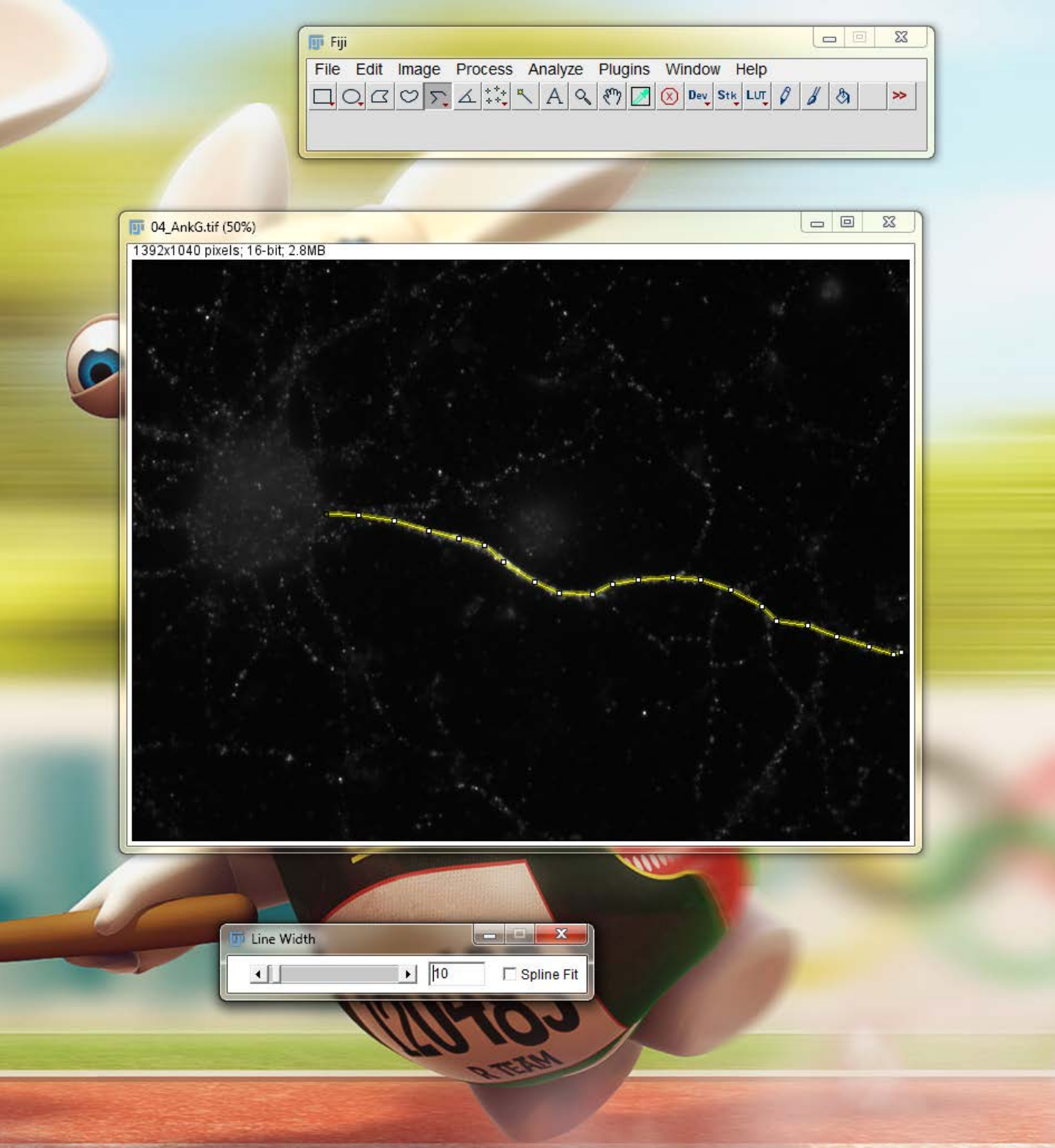




- Open the segmented line tool (right click line tool) and draw a line over the region of interest.
- The region of interest is in this case the axon starting from the soma.

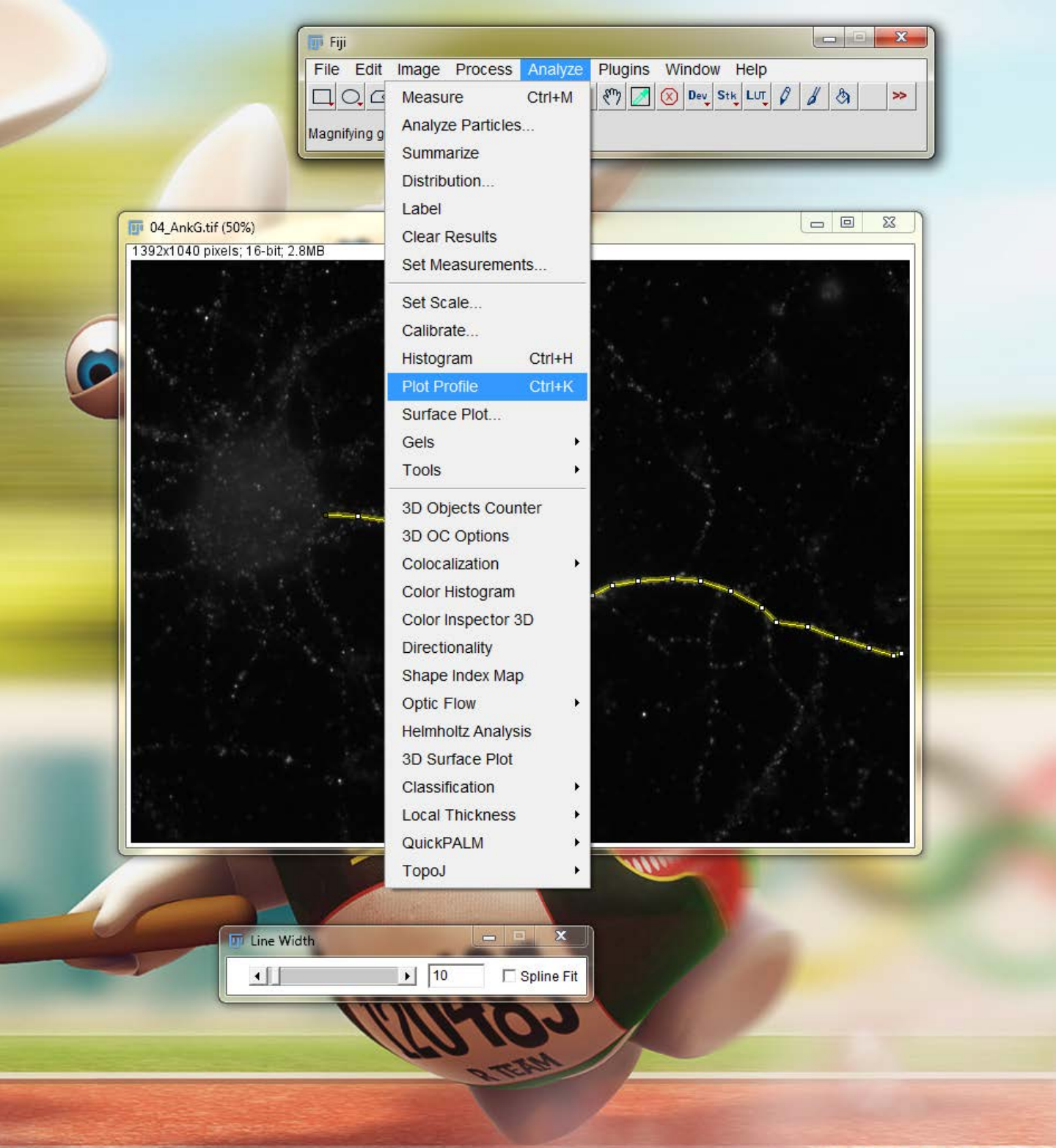


- Like this...

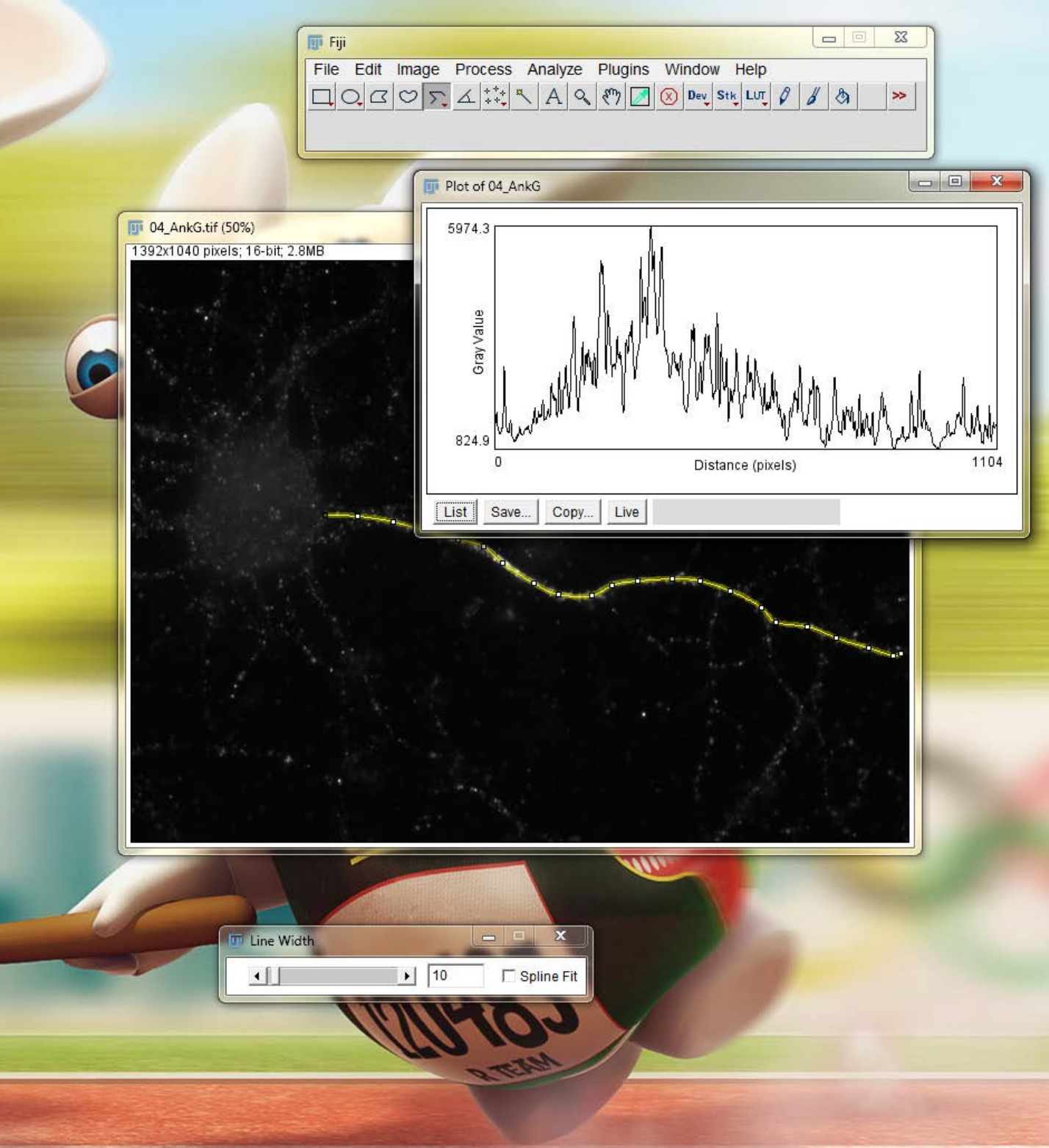


- You can make the line thicker by adjusting the line width (double click the segmented line tool to open line width window)

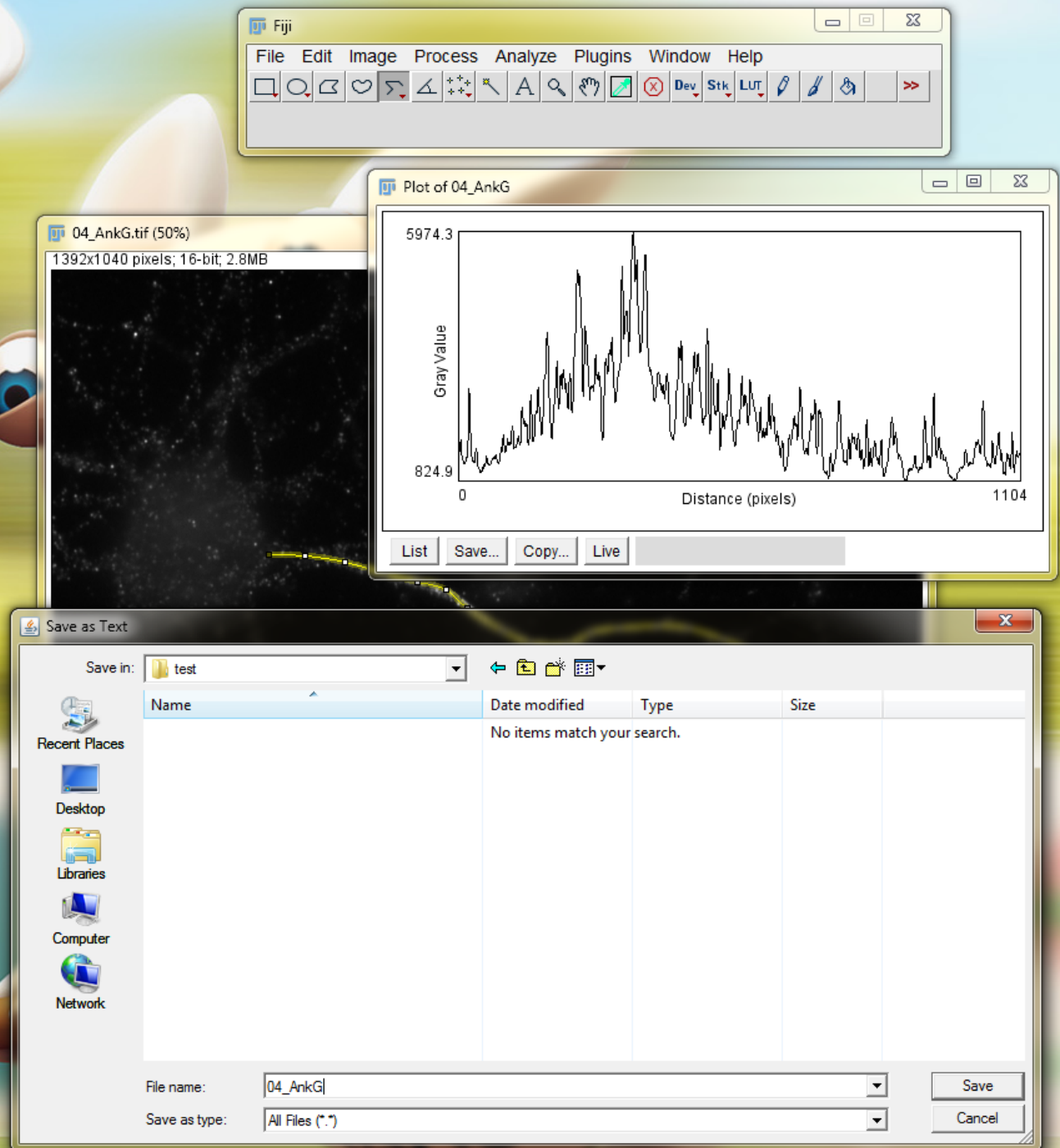




- Plot the profile of the line

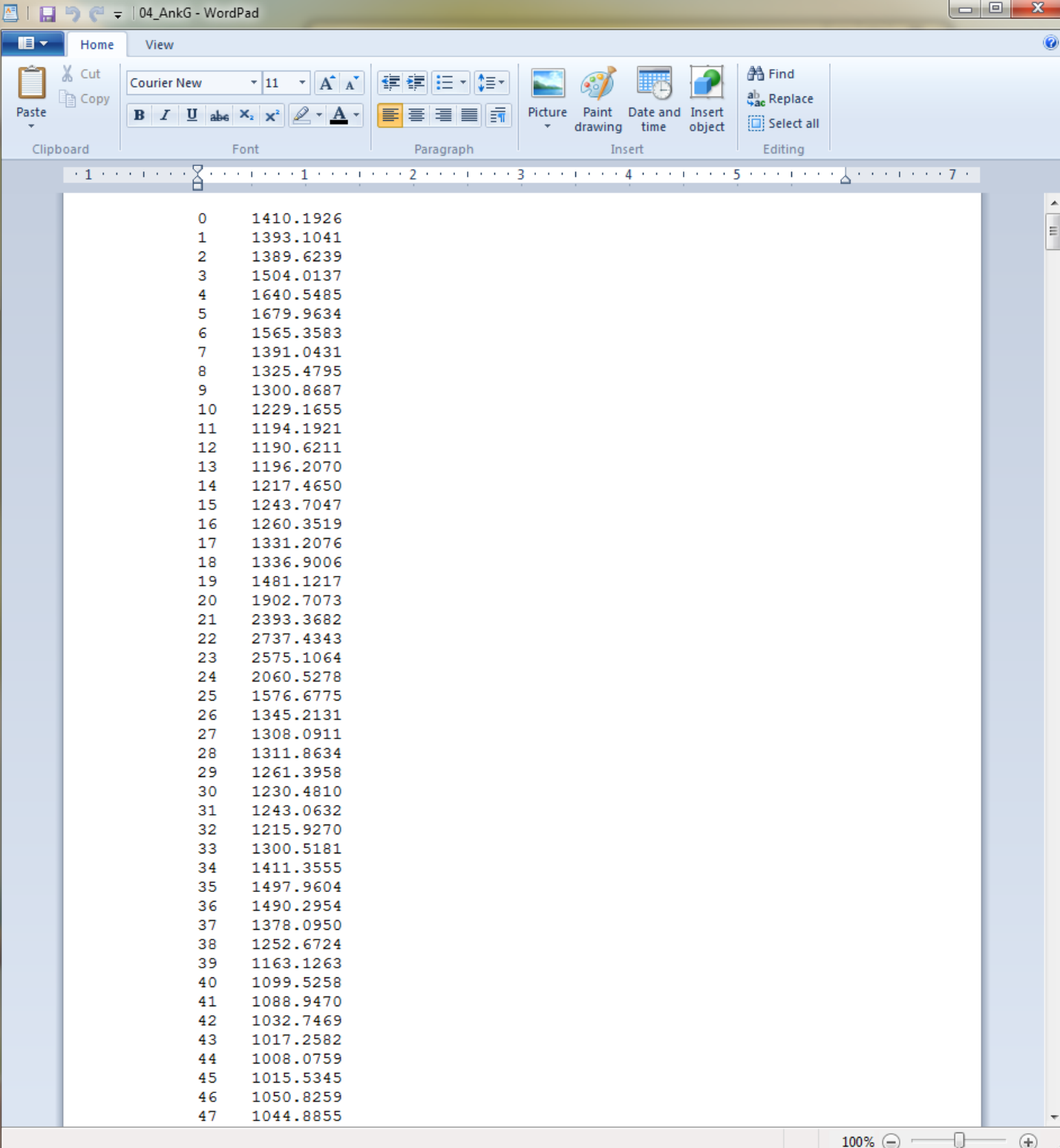


- It should look something like this....



- Click save and save the profile as a .txt file in a folder where you will save all the profiles of a specific staining.
- It is easier to start with a number followed by the staining (this helps to organize your results)

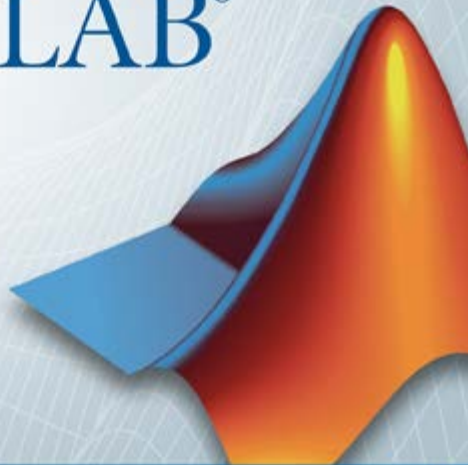




- The .txt file will look something like this...
- In the first column you will find the distance in pixels (or in microns if you did not reset the scale to pixels).
- In the second column you will find the intensity of the line.

# MATLAB®

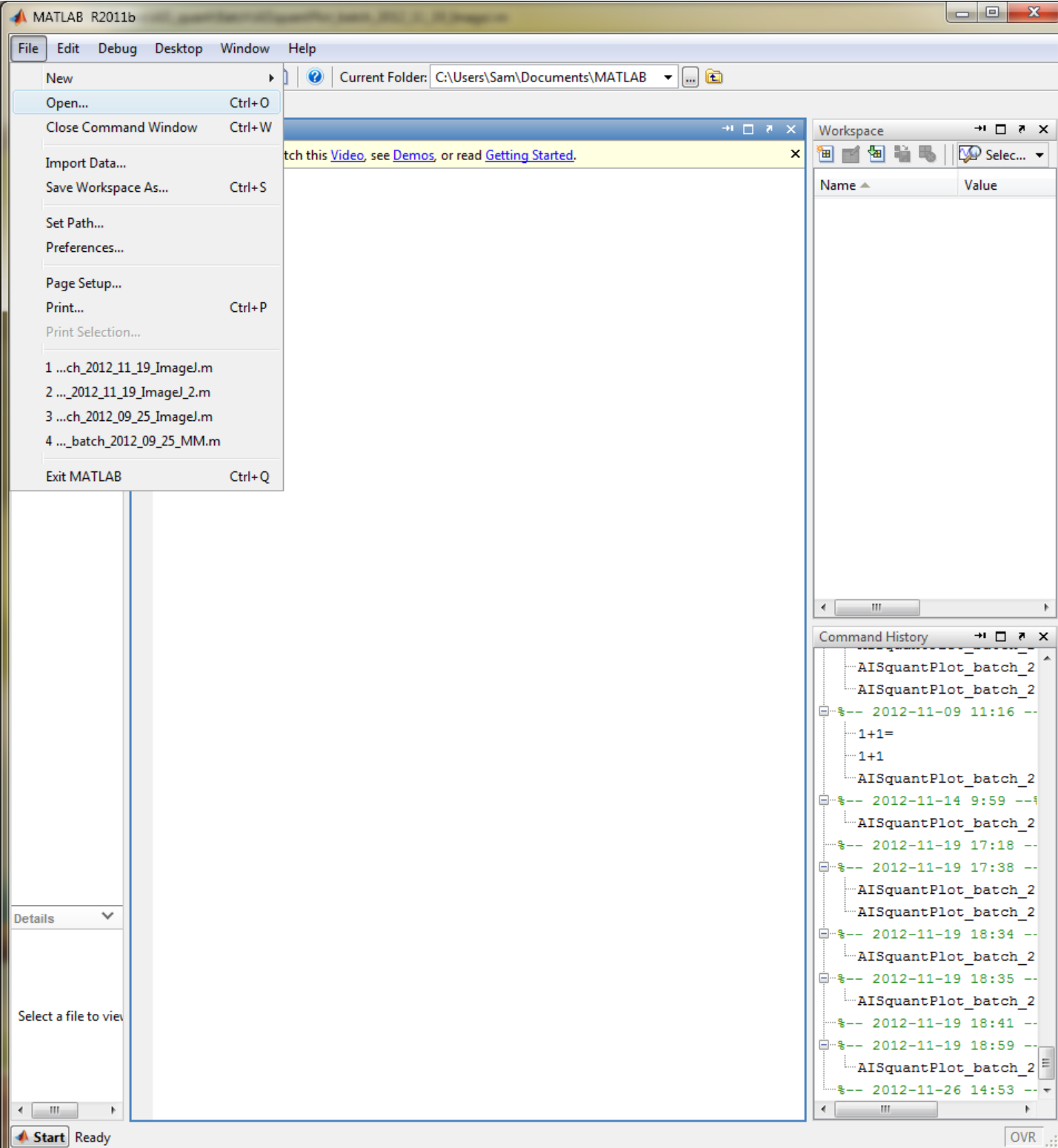
R2011b (7.13.0.564)  
64-bit (win64)  
August 14, 2011  
License Number: 3254



R2011b

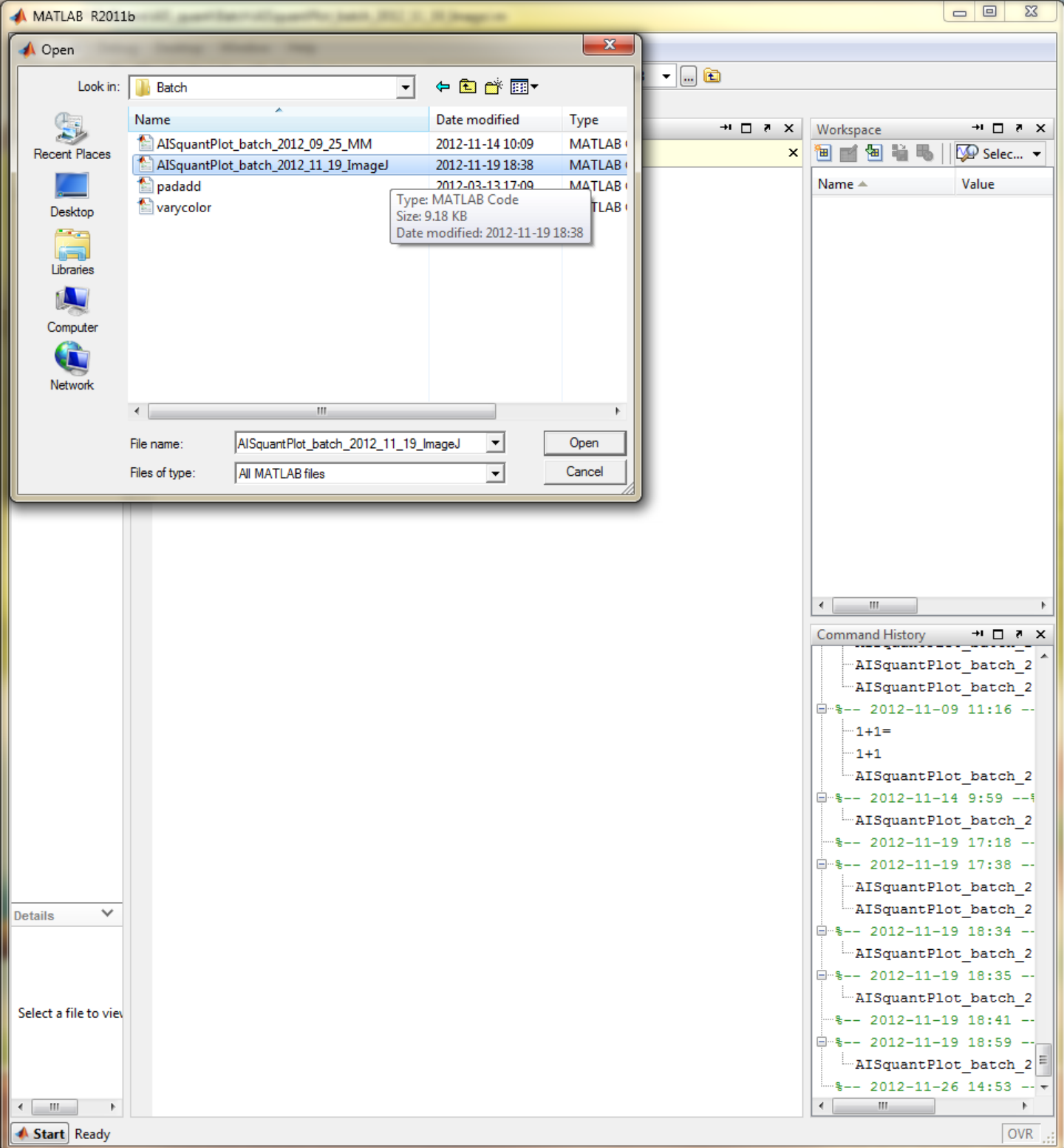
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- Open Matlab



- Open the AIS\_quant script



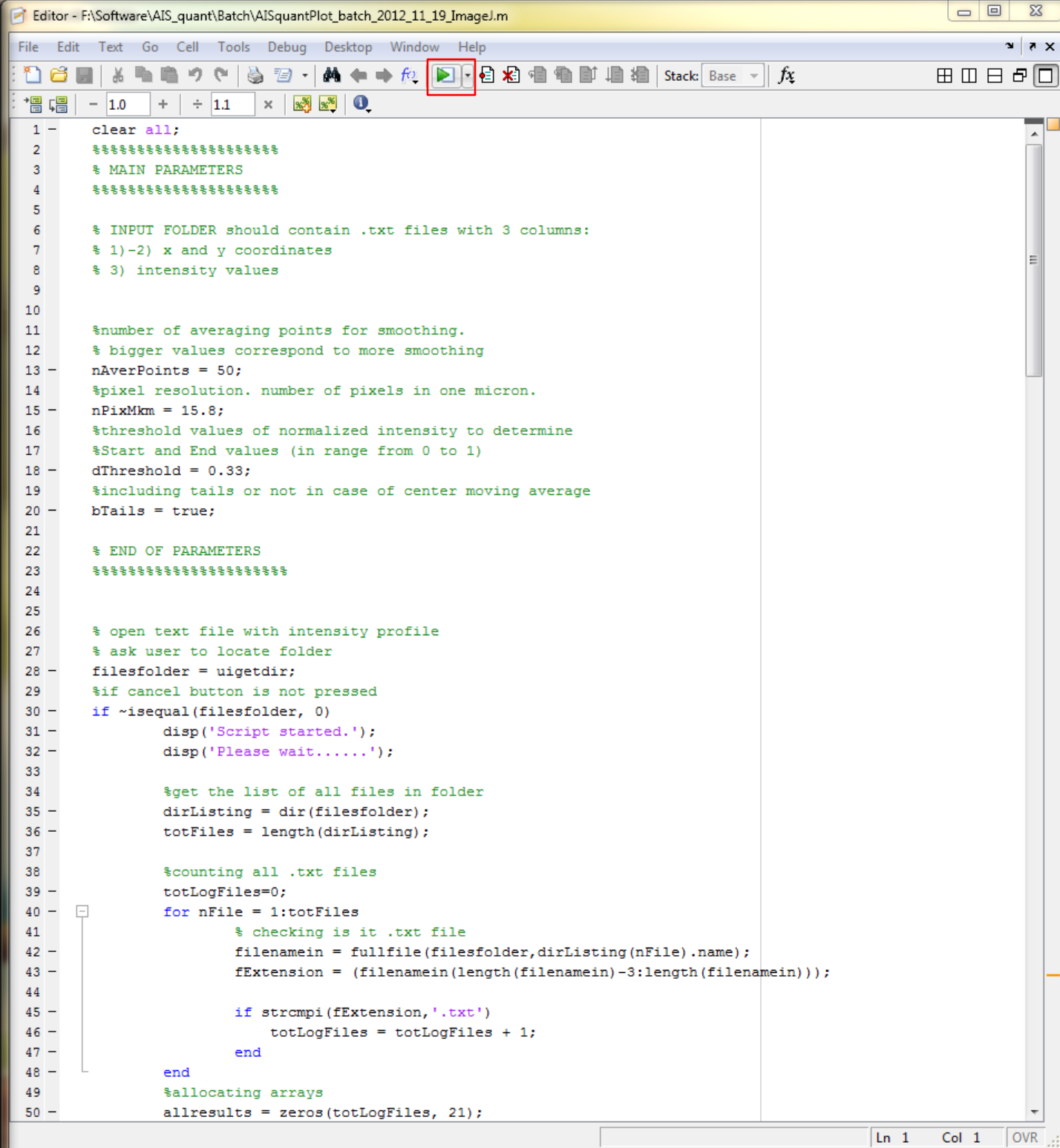


- Be sure that you open the ImageJ version

```
Editor - F:\Software\AIS_quant\Batch\AISquantPlot_batch_2012_11_19_ImageJ.m
File Edit Text Go Cell Tools Debug Desktop Window Help
[Icons] [fx] Stack: Base [fx] [Icons]
- 1.0 + ÷ 1.1 × [Icons] [?]
1 - clear all;
2 - %%%%%%%%%%%%%%%%%%%%%%%%%
3 - % MAIN PARAMETERS
4 - %%%%%%%%%%%%%%%%%%%%%%%%%
5 -
6 - % INPUT FOLDER should contain .txt files with 3 columns:
7 - % 1)-2) x and y coordinates
8 - % 3) intensity values
9 -
10 -
11 - %number of averaging points for smoothing.
12 - % bigger values correspond to more smoothing
13 - nAverPoints = 50;
14 - %pixel resolution. number of pixels in one micron.
15 - nPixMkm = 15.8;
16 - %threshold values of normalized intensity to determine
17 - %Start and End values (in range from 0 to 1)
18 - dThreshold = 0.33;
19 - %including tails or not in case of center moving average
20 - bTails = true;
21 -
22 - % END OF PARAMETERS
23 - %%%%%%%%%%%%%%%%%%%%%%%%%
24 -
25 -
26 - % open text file with intensity profile
27 - % ask user to locate folder
28 - filesfolder = uigetdir;
29 - %if cancel button is not pressed
30 - if ~isequal(filesfolder, 0)
31 -     disp('Script started. ');
32 -     disp('Please wait.....');
33 -
34 -     %get the list of all files in folder
35 -     dirListing = dir(filesfolder);
36 -     totFiles = length(dirListing);
37 -
38 -     %counting all .txt files
39 -     totLogFiles=0;
40 -     for nFile = 1:totFiles
41 -         % checking is it .txt file
42 -         filenamein = fullfile(filesfolder, dirListing(nFile).name);
43 -         fExtension = (filenamein(length(filenamein)-3:length(filenamein)));
44 -
45 -         if strcmpi(fExtension, '.txt')
46 -             totLogFiles = totLogFiles + 1;
47 -         end
48 -     end
49 -     %allocating arrays
50 -     allresults = zeros(totLogFiles, 21);
```

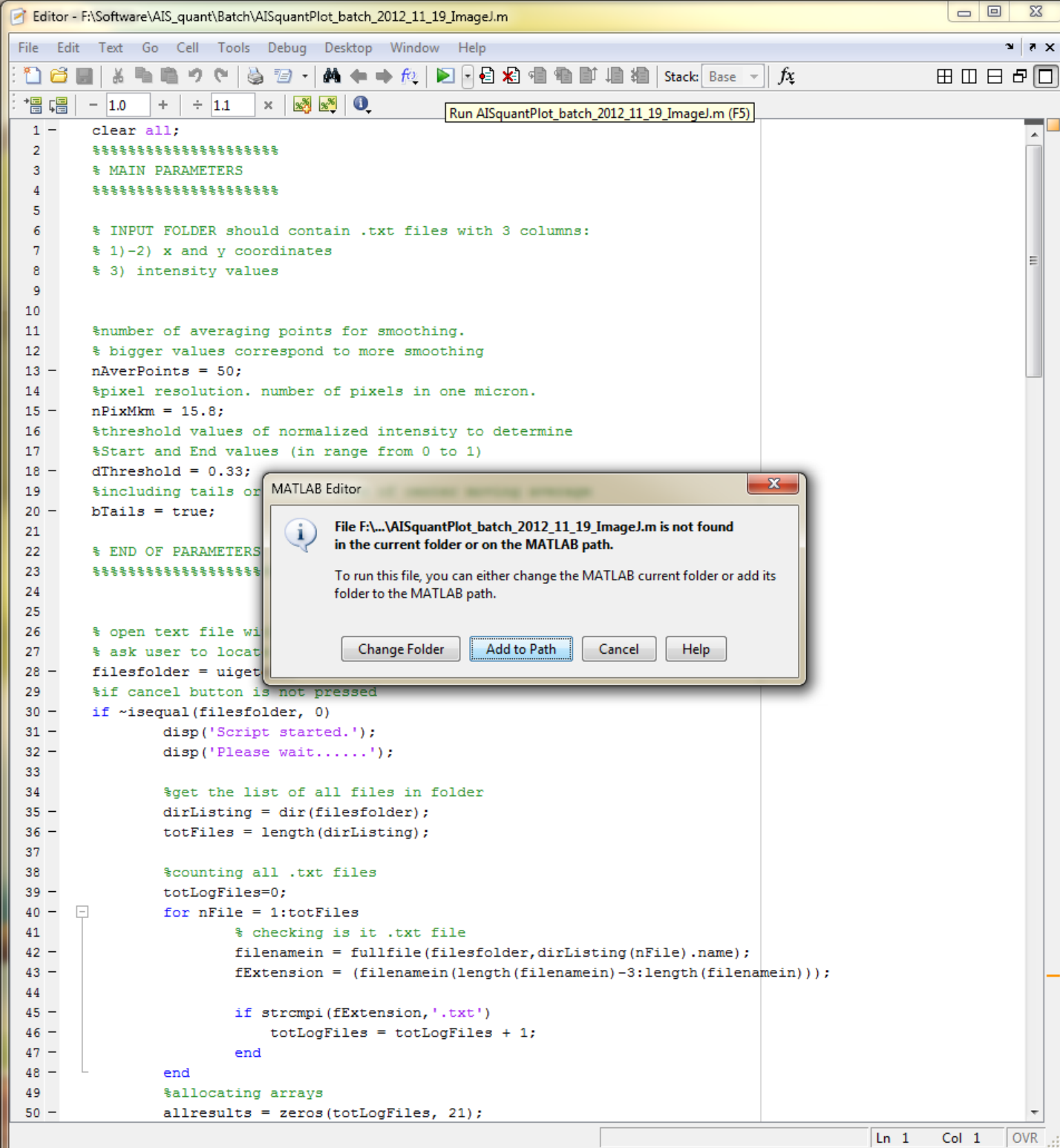
Ln 1 Col 1 OVR

- This is how the script looks like
- There are three important parameters (red box)
- AverPoints: the point that are used for smoothing (I always use the amount of pixels equal to  $\sim 3\mu\text{m}$ )
- PixMkm: number of pixels in one micron of the picture (use 1 here when ImageJ already used the correct scale)
- Treshold: treshold value (range 0-1) to determine to start and end values

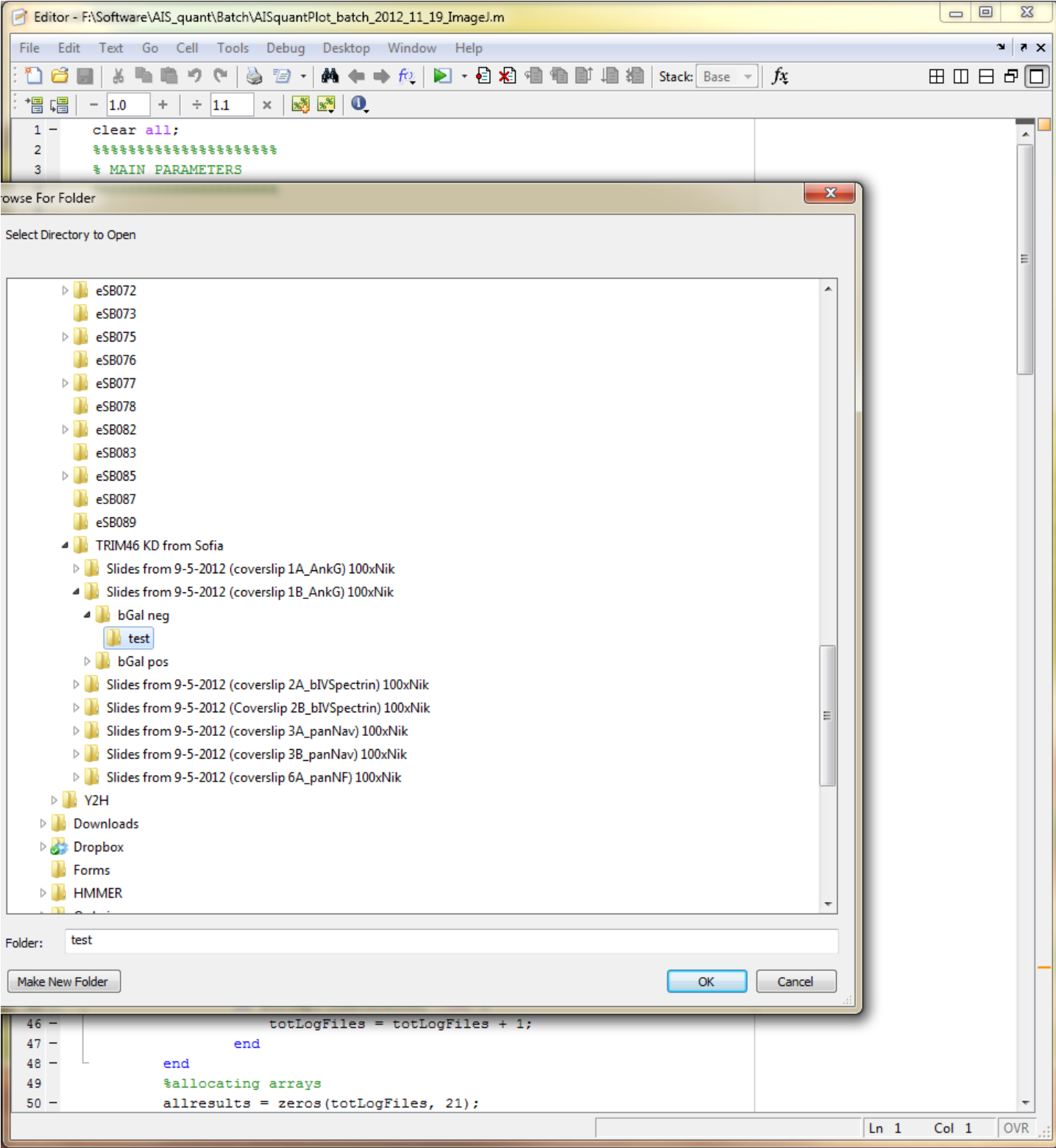


- Run the script using the button shown by the red square





- Click Add to Path



- Locate the folder with your plot data from ImageJ (.txt)
- Click OK

MATLAB R2011b

File Edit Debug Desktop Window Help

Current Folder: C:\Users\Sam\Documents\MATLAB

Shortcuts How to Add What's New

Command Window

New to MATLAB? Watch this [Video](#), see [Demos](#), or read [Getting Started](#).

```
>> AISquantPlot_batch_2012_11_19_ImageJ
Script started.
Please wait.....
04_AnkG.txt finished
Analysis done. Saving...
Done.
fx >> |
```

Workspace

Name	Value
End1Ind	582
End2Ind	523
EndVal1	36.7722
EndVal2	33.0380
LengthVal1	27.3418
LengthVal2	23.6076
LengthVal3	27.3418
LengthVal4	23.6076
MaxInd	347
MaxVal	21.8987
StartInd1	150
StartInd2	150
StartVal1	9.4304
StartVal2	9.4304
allnormtrace	<1105x2 dou
allresults	<1x21 doubl
bLeftTail1	2.4250e+04
bLeftTail2	2.4250e+04
bMeanInt11	4.6628e+04
bMeanInt12	4.8208e+04
bMeanInt21	4.6628e+04

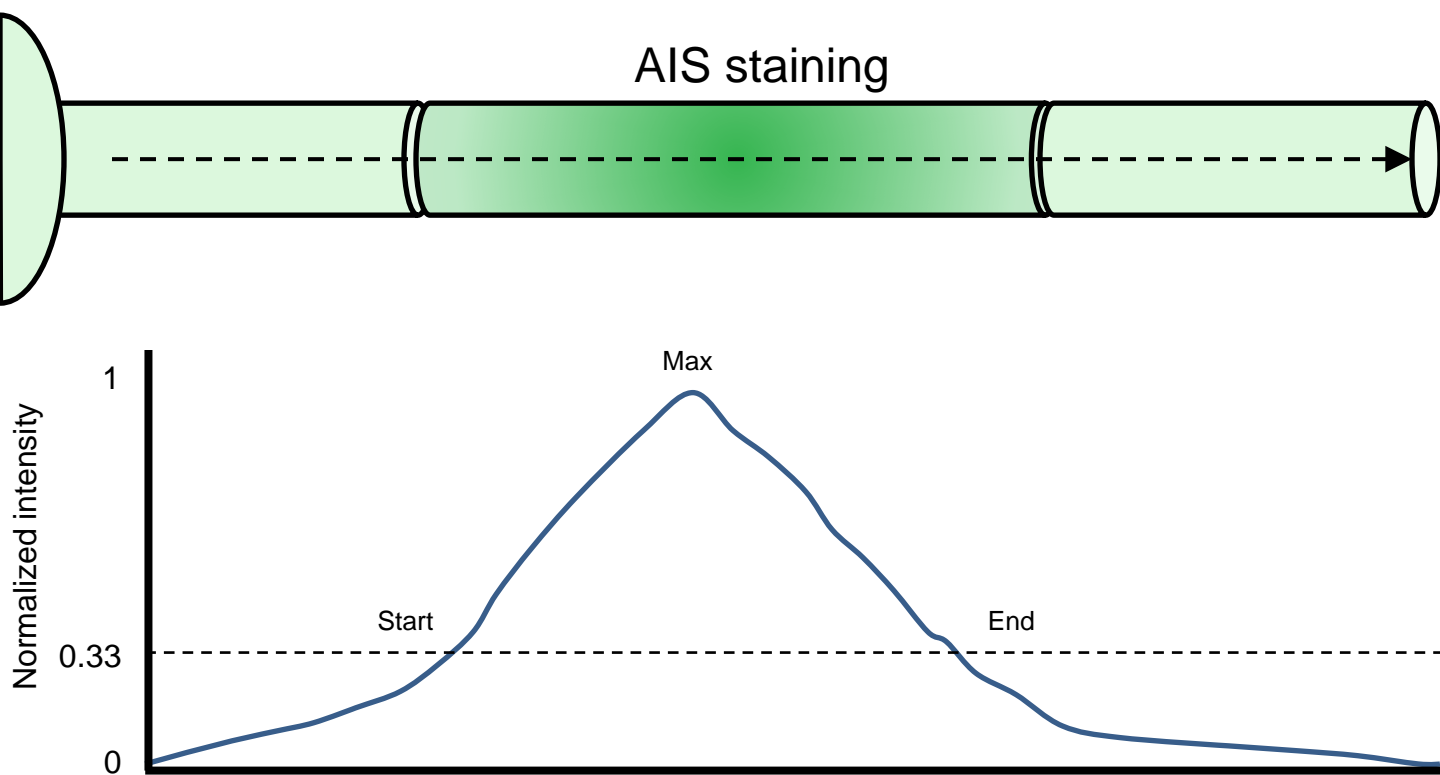
Command History

```
1+1=
1+1
AISquantPlot_batch_2
$-- 2012-11-14 9:59 --$
AISquantPlot_batch_2
$-- 2012-11-19 17:18 --$
$-- 2012-11-19 17:38 --$
AISquantPlot_batch_2
AISquantPlot_batch_2
$-- 2012-11-19 18:34 --$
AISquantPlot_batch_2
$-- 2012-11-19 18:35 --$
AISquantPlot_batch_2
$-- 2012-11-19 18:41 --$
$-- 2012-11-19 18:59 --$
AISquantPlot_batch_2
$-- 2012-11-26 14:53 --$
AISquantPlot_batch_2
$-- 2012-11-26 14:57 --$
AISquantPlot_batch_2
```

Start

OVR

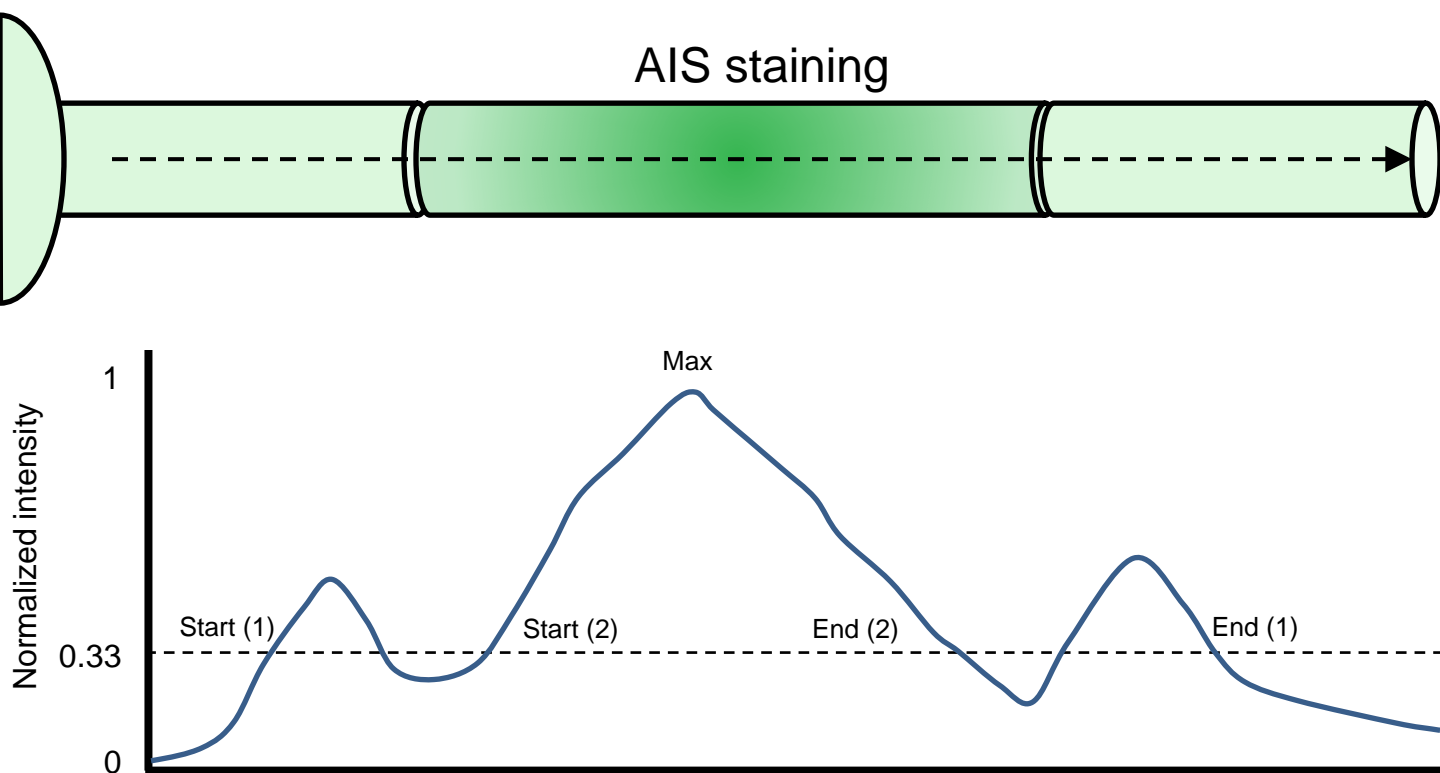
- The script will automatically analyze all .txt files located in the folder and save the results in a single Excel file (named summary)



But what was actually analyzed by the script?

- It converts pixels to  $\mu\text{m}$  (mkm).
- It automatically normalizes all data from 0 (lowest value) to 1 (highest data).
- It determines the max value of the increased staining, which is equal to the position where the normalized intensity is 1.
- It determines the start and end value of the increased staining, which is equal to the positions where the normalized intensity crosses the threshold.





But what if the threshold is crossed more than twice?

The start is determined by:

- Start(1): from the first data point (increase from the first data point to the threshold)
- Start(2): from the max (decrease from Max to the threshold)

The end is determined by:

- End(1): from the last data point (increase from the last data point to the threshold)
- End(2): from the max (decrease from Max to the threshold)

All start and end values are saved by the script in the Excel file

summary [Compatibility Mode] - Microsoft Excel

	A	B	C	D	E	F	G	H	I	J	K	L	M
1	File:	04_AnkG.txt											
2	length, mkm	normalized intensity											
3	0	0.135429717											
4	0.063291139	0.133172678											
5	0.126582278	0.130698462											
6	0.189873418	0.128432007											
7	0.253164557	0.125836513											
8	0.316455696	0.123123843											
9	0.379746835	0.120692938											
10	0.443037975	0.118174662											
11	0.506329114	0.116514619											
12	0.569620253	0.115853273											
13	0.632911392	0.115915283											
14	0.696202532	0.115914814											
15	0.759493671	0.11507165											
16	0.82278481	0.113353849											
17	0.886075949	0.111083											
18	0.949367089	0.108480184											
19	1.012658228	0.105929423											
20	1.075949367	0.103124274											
21	1.139240506	0.100346163											
22	1.202531646	0.097633284											
23	1.265822785	0.095084635											
24	1.329113924	0.092858749											
25	1.392405063	0.090690286											
26	1.455696203	0.088755105											
27	1.518987342	0.087214912											
28	1.582278481	0.085628691											
29	1.64556962	0.084301128											
30	1.708860759	0.082983297											
31	1.772151899	0.081970599											
32	1.835443038	0.080559538											
33	1.898734177	0.078714199											
34	1.962025316	0.076820072											
35	2.025316456	0.075145987											
36	2.088607595	0.074113937											
37	2.151898734	0.073227063											
38	2.215189873	0.072600284											
39	2.278481013	0.072402789											
40	2.341772152	0.072243995											

In sheet 2 of the Excel file you will find the distance in mkm ( $\mu\text{m}$ ) and the normalized intensity corresponding to the distance.

summary [Compatibility Mode] - Microsoft Excel

	A	B	C	D	E	F	
1	Filename	Start from left (1), mkm	Start from max value(2), mkm	Max, mkm	End from right end (1), mkm	End from max value (2), mkm	AI Sler
2	04_AnkG.txt	9.430379747	9.430379747	21.89873418	36.7721519	33.03797468	
3							

summary [Compatibility Mode] - Microsoft Excel

	G	H	I	J	K	L
1	AI Slength (End1-Start1), mkm	AI Slength (End2-Start1), mkm	AI Slength (End1-Start2), mkm	AI Slength (End2-Start2), mkm	# of smooth points	intensity thres
2	27.34177215	23.60759494	27.34177215	23.60759494	51	
3						

summary [Compatibility Mode] - Microsoft Excel

	L	M	N	O	P	Q	R
1	intensity threshold	resolution (pixels in mkm)	Tails	Int_Left_tail1, a.u./mkm	Int_Left_tail2, a.u./mkm	Int_Middle_(Start1_End1)	Int_Middle_(Start2_End1)
2	0.33	15.8	1	24250.19154	24250.19154	46627.64844	46627.64844
3							

summary [Compatibility Mode] - Microsoft Excel

	S	T	U	V	W	X	Y	Z	AA
1	Int_Middle_(Start1_End2)	Int_Middle_(Start2_End2)	Int_Right_tail1, a.u./mkm	Int_Right_tail2, a.u./mkm					
2	48208.41151	48208.41151	23524.20319	24853.2043					
3									
4									

In sheet 1 of the Excel file you will find:

- All the different start and end values as well as the max value.
- The length, which is the difference between the different start and end values.
- The parameters you used for the script.
- The mean intensities of the raw dataset between the different start and end values.
- The mean intensities of the raw dataset of the different tails, either before the start or after the end value.



You can now use all this data to determine the position on your staining using, for instance, different conditions.

Are some things unclear?

Try to read this manual again...

Still unclear?

Really?

OK you can always drop by or send me an email with your question....

Cheers,  
Sam  
([s.vanbeuningen@uu.nl](mailto:s.vanbeuningen@uu.nl))